Molecular Detection of *Vibrio* spp. in Fish and Shrimp from the Persian Gulf

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ABSTRACT: *Vibrio* species are the major seafood-borne bacteria that are frequently associated with the consumption of contaminated sea food. A total of 113 samples including 58 fish and 55 shrimps were studied for the possible contamination with *Vibrio* species. A biochemical protocol was applied for the identification of the *Vibrio* isolates and Polymerase Chain Reaction (PCR) was carried out to confirm the strains. The results indicated that 25 samples (22%) were contaminated with *Vibrio* species. Among *Vibrio* isolates, *Vibrio harveyi* was the species most frequently isolated (11.5%), followed by followed by *V. parahaemolyticus*, *V. vulnificus*, *V. alginolyticus* and *V. mimicus*. *Vibrio cholerae* was not detected in the studied samples. The results of this study indicated that fish and shrimp from the Persian Gulf regularly contain pathogens that might affect the public health.

Keywords: Fish, PCR, Persian Gulf, Shrimp, *Vibrio* spp.

Introduction

Members of the genus *Vibrio* are motile, Gram-negative straight or curved rods (Roberts, 2001). They are facultative anaerobic chemo-organotrophs capable of both respiratory and fermentative metabolism (Roberts, 2001; Raissy et al., 2012). Some species, such as *V. parahaemolyticus*, *V. vulnificus* and *V. cholerae* are regularly linked to human food-borne infections caused by the consumption of raw, undercooked contaminated sea foods, but there are occasional reports of food-borne or waterborne infections caused by the environmental *Vibrio* or *Vibrio*-like species (Messelhausser et al., 2012). Other *Vibrio* species consist of *V. alginolyticus*, *V. harveyi*, *V. fluvialis*, *V. furnissii*, *V. metschnikovii*, *V. mimicus* and *V. parahaemolyticus* that might cause disease in both aquatic animals and humans (Austin, 2012). Pathogenic *Vibrio* species such as *V. cholerae*, *V. parahaemolyticus*, and *V. vulnificus* cause gastrointestinal illnesses or septicemia that might lead to fatal complications (Faruque and Nair, 2006; Janda et al., 1988). The mortality rate of *V. vulnificus* septicemia exceeds 50% and approaches 100% in cases of septic shock (Kumamoto & Vukich, 1998). Seafood products harvested from contaminated water or have been improperly preserved after harvesting are known to play an important role in the infections by bacteria (Mouzin et al., 1997). Occurrence of *Vibrio* in fish and shellfish has been reported in different countries including Iran (Janda et al., 1988; Jaksic et al., 2002; Hosseini et al., 2004; Rahimi et al., 2010) while the epidemiology of vibriosis, except *Vibrio cholera*, is essentially unknown in Iran. In this study the occurrence of *vibrio* spp. in fish and shrimp from the Persian Gulf and the possible harms for human is studied.

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Materials and Methods

- **Sample preparation**
  A total number of 113 samples including 58 fish and 55 shrimps were collected from the local fish market during August 2010 to April 2011. The samples were transferred into cool boxes with an internal temperature of +2 to +4°C after collection and were processed within a short time after arrival.

- **Bacteriological Analysis**
  The *Vibrio* spp. analysis took place according to the method described by Bockemühl (1992). 25 g of the homogenized meat was added to 225 ml of alkaline peptone water (APW) and incubated at 37°C. The samples were subcultivated on Thiosulfate Citrate Bile Salts Sucrose agar (TCBS, BD Diagnostics, Heidelberg, Germany) and on modified Cellobiose Polymyxin-B Colistin agar (mCPC). After the incubation at 37°C for 24h, the isolates were used for further screening tests including gram staining, oxidase and catalase tests, culture in SIM and TSI media and other biochemical tests as described by Hosseini *et al.* (2004).

- **DNA Extraction**
  The genomic DNA was prepared using a standard DNA extraction method as described by Ausubel *et al.* (1987). The purity and quantity of genomic DNA in each sample was evaluated by measuring the optical densities at 260 and 280nm wavelength. The DNA concentration of each sample was adjusted to 50ng/µl for PCR.

- **PCR assay**
  Two sets of oligonucleotide primers were used for specific identification of each *Vibrio* species. The primer sequences, targeting genes and amplicon sizes are listed in Table 1. The PCR reaction was performed in a 50 µl reaction system consisting of 2 µl of purified genomic DNA (50 ng/µl), 5 µl of 10×PCR buffer (100 mM Tris–HCl, pH 8.3, 500 mM KCl, 60 mM MgCl2, 0.1% gelatin and 1% Triton X-100), 1 µl each of the primers (50 pmol/µl), 1 µl each of the 10 mM dNTPs, 0.2 µl units Taq DNA polymerase (5 units/µl) and 40 µl of sterile distilled water. The reactions were performed with a PTC-100 thermal cycler (Eppendorf, Harburg, Germany) with thermal cycling profile as is indicated in Table 1. Amplified products were separated by electrophoresis in ethidium bromide stained with 1.5% agarose gels at 90 V for 50 min. The gels were visualized and photographed with a UV transilluminator.

Results and Discussion

A total number of 113 samples consisting of 58 fish and 55 shrimps were analysed for *Vibrio* spp. using both biochemical tests and PCR. The results revealed that 25 samples (22.1%) including 18 fish and 7 shrimps contained *Vibrio* species. A total of 25 isolates were identified in this study where the most frequent species were *Vibrio* *harveyi* followed by *V. parahaemolyticus*, *V. vulnificus*, *V. alginolyticus* and *V. mimicus*. The number of samples contained *Vibrio* species are presented in Table 2.

Many studies show the presence of *Vibrio* species in aquatic animals such as fish (Messelhausser *et al.*, 2012; Schmidt *et al.*, 2000), shrimp (Raissy *et al.*, 2012; Rahimi *et al.*, 2012; Reboucas *et al.*, 2011; Lhafi and Kuhne, 2009), lobster and crab (Raissy *et al.*, 2012) and mussel (Lhafi and Kuhne, 2009).

In the present study, five *Vibrio* species including *V. vulnificus*, *V. parahaemolyticus*, *V. alginolyticus*, *V. harveyi* and *V. mimicus* were detected from the examined samples which is in agreement with the results of previous studies (Jaksic *et al.*, 2002; Hosseini *et al.*, 2004; Rahimi *et al.*, 2010, Lhafi and Kuhne, 2009). According to the results, 29.3 % of the fish and 12.7 % of the shrimps samples examined revealed the presence of *Vibrio* species.

Among the *Vibrio* species detected in this
Table 1. Primer sequences, targeting genes and amplicon size of primers

<table>
<thead>
<tr>
<th>Target species</th>
<th>Sequence(5'------ 3')</th>
<th>PCR program*</th>
<th>Targeting Gene</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>V. parahaemolyticus</td>
<td>GCAGCTGATCAAAACGTGTAAGTATTATCGTATCGTCACCTAC</td>
<td>a</td>
<td>flaE</td>
<td>(Tarr et al., 2007)</td>
</tr>
<tr>
<td>V. cholerae</td>
<td>AAGACCTCAACTGGCGGTA</td>
<td>b</td>
<td>sodB</td>
<td>(Tarr et al., 2007)</td>
</tr>
<tr>
<td>V. vulnificus</td>
<td>GCCTCAAGCTGGTGAAGGAAGTGTTAGTGATTGCTAGAG</td>
<td>c</td>
<td>hsp</td>
<td>(Tarr et al., 2007)</td>
</tr>
<tr>
<td>V. mimicus</td>
<td>CATTCCGGTTCTTTCGCTGAT</td>
<td>d</td>
<td>sodB</td>
<td>(Tarr et al., 2007)</td>
</tr>
<tr>
<td>V. alginolyticus</td>
<td>CGAGTACAGTCACTTGAAAGCCACAACAGAACTCGCGTTACC</td>
<td>e</td>
<td>collagenase</td>
<td>(Di Pinto et al., 2005)</td>
</tr>
<tr>
<td>V. harveyi</td>
<td>CTTCACGCTTGATGGCTACTG</td>
<td>f</td>
<td>vhh</td>
<td>(Maiti et al., 2009)</td>
</tr>
</tbody>
</table>

* PCR program: a, b, c, d: 35 times (92°C, 40 s; 57°C, 1 min; 72°C, 1.5 min); e: 35 times (94°C, 30 s; 57°C, 30 sec; 72°C, 1 min); f: 30 times (95°C, 1 min; 50°C, 1 min; 72°C, 1 min)

Table 2. The number of samples containing vibrio species

<table>
<thead>
<tr>
<th>Sample</th>
<th>V. vulnificus</th>
<th>V. parahaemolyticus</th>
<th>V. mimicus</th>
<th>V. alginolyticus</th>
<th>V. harveyi</th>
</tr>
</thead>
<tbody>
<tr>
<td>Scomberomorus commerson (24)</td>
<td>0</td>
<td>2</td>
<td>0</td>
<td>1</td>
<td>0</td>
</tr>
<tr>
<td>Otolithes ruber (15)</td>
<td>2</td>
<td>1</td>
<td>1</td>
<td>0</td>
<td>5</td>
</tr>
<tr>
<td>Scomberomorus guttatus (14)</td>
<td>1</td>
<td>0</td>
<td>0</td>
<td>1</td>
<td>3</td>
</tr>
<tr>
<td>Acanthopagrus latus (5)</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>1</td>
<td>0</td>
</tr>
<tr>
<td>penaeus semisulcatus (55)</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>1</td>
<td>5</td>
</tr>
<tr>
<td>Total (113)</td>
<td>3</td>
<td>4</td>
<td>2</td>
<td>3</td>
<td>13</td>
</tr>
</tbody>
</table>

study, V. parahaemolyticus has often been reported to cause gastro-intestinal problems following the consumption of contaminated seafood. In Japan, it has been reported to be responsible for one-fourth of all gastro-intestinal cases caused by food while in the USA, it has caused 14 outbreaks of food poisoning between 1971 and 1978 (Feldhusen, 2000). The occurrence of this species in Iran has been reported in shrimp, lobster and crab from the Persian Gulf (Raissy et al., 2012; Hosseini et al., 2004; Rahimi et al., 2010). The contamination of shrimp from the Persian Gulf with this species was reported in the order of 9.3% by Rahimi et al. (2010). Hosseini et al. (2004) reported four isolates of V. parahaemolyticus in 770 studied shrimp samples from the same place. In the present study, four samples (2.6%) were found to be contaminated with V. parahaemolyticus. Differences in contamination of seafood with Vibrio species might be related to the catching method, transportation condition, time of examination as well as seasonal and environmental conditions.

As an important human pathogenic bacterium, V. vulnificus has been associated with a small but increasing number of serious life-threatening conditions such as gastro-enteritis and wound infections which might become septicaemic (Mouzopoulos et al., 2008). The onset of symptoms is often abrupt, with a rapid progression to septic shock and thus death despite the intervention of antibiotics (Haq and Dayal, 2005). In the USA, V. vulnificus has been regarded as being responsible for most of the seafood related deaths since the first report in 1979 (Oliver, 2005). Indeed, a regular source of infection with this pathogen is the consumption of contaminated raw or under-cooked seafood (Drake et al., 2007).

V. harveyi is proved to be a serious pathogen of shrimp and fish causing luminous bacterial disease in penaeid shrimp (Austin, 2010). In terms of human disease, V. harveyi has been recovered from wound infections, specifically from a leg wound resulting from a shark bite in USA (Pavia et al., 1989).

Conclusion

The results of this study indicated that vibrio species is a potential pathogen that
might be found in fish and shrimps caught in the Persian Gulf and might put human health at risk if consumed. The bacteria will be removed by using high cooking temperature, although the toxin might remain in the food stuff depending on the processing conditions.

References


